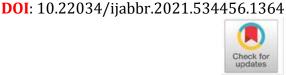
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Original Article



Screening of bacteria *Streptomyces* Waksman and Henrici 1943 (Streptomycetaceae) Isolates from Soil Samples in Iraq

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ABSTRACT

Background: The genus *Streptomyces* Waksman & Henrici 1943 includes aerobic, grampositive, and filamentous bacteria which produce well developed vegetative hyphae with branches. The wall consists of a large mixture of different compounds, including peptidoglycan, teichuronic acid, teichoic, and polysaccharides. The peptidoglycan components consist of glycan as a chains of irregular N-acetyl- d-muramic acid (NAM), diaminopimelic acid, and N-acetyl-d-glucosamine (NAG) and DAP, which is unique in the cell walls of prokaryotic microorganisms. The teichoic and teichuronic acid are chemically bonded to peptidoglycan.

Methods: One gram of soil samples was used to make suspension, by adding 99 mL of sterile distilled water (stock suspension) into it and shaking it in a shaker at 160 rpm for 30 minutes at room temperature. Serial dilutions from 0.1-0.001 were made from the stock suspension, and left for 10 minutes. After shaking, 0.1 mL of each dilution was cultured on Yeast Extract and Malt Extract agar (YEME) with Streptomycin 50 ug/mL. The inoculated plates were incubated at 28 °C for 7 to 10 days. Based on cultural characteristics, suspected colonies of *Streptomyces* were selected, which are characterized as small, white, pin-point, rough, chalky, and a clear zone of inhibition around them. These colonies were confirmed their identification by types of Gram's stain, aerial and substrate mycelium color, pigment production, and pigment color. *Streptomyces* were re-streaked on International *Streptomyces* project (ISP) to obtain pure colonies used for identification.

Results: The current study aimed to screen bacteria *Streptomyces* isolates. Only 21 samples of soil were suspected to contain *Streptomyces*, and 45 isolates were obtained with different morphology types per samples of soil. The colonies suspects were selected basis on color that ranged from gray, white and creamy. The microscopic examination of local *Streptomyces* spp. after Gram-staining method was conducted. The observations revealed that local *Streptomyces* is gram positive and rod shaped similar to

features of fungal in possessing branched mycelium. The *Streptomyces* produced extra cellular enzymes like amylase, urease, catalase, protease, Gelatinase, cellulase and phosphatase. Utilization of citrate was positive, with no Melanine reaction production and soluble pigmented, and negative for indole production.

Conclusion: The identification of the *Streptomyces* is a very complex process. Morphological and biochemical characteristics are two important aspects for the classification in the Streptomycetaceae family. By studying the morphological, cultural, and biochemical characteristics, it is observed that the local isolates are belonging to the genus of *Streptomyces*.

Keywords: Kidney Stones, Vitamin D, Hypercalciuria.

1. Introduction

Streptomyces Waksman & Henrici 1943 is the type genus of the family Streptomycetaceae [1] and currently covers close to 576 species with the number increasing every year [2, 3]. Streptomyces includes aerobic, Grampositive, and filamentous bacteria which produce well developed vegetative hyphae (between 0.5-2.0 μm in diameter) with branches. They form a complex substrate mycelium that scavenging organic compounds from their substrates [4]. Although mycelium and the aerial hyphae that arise from them are amotile, mobility is achieved by dispersion of spores [4]. Spore surfaces may be hairy, rugose, smooth, spiny or warty [5].

The actinomycetes cell wall is an inflexible structure that maintains the cell wall of actinomycetes shape through cell wall which prevents bursting of the cell high osmotic pressure [6]. The wall consists of a large mixture of different compounds, including peptidoglycan, teichuronic acid, teichoic, and polysaccharides. peptidoglycan The components consists of glycan as a chains of irregular N-acetyl-d-muramic acid (NAM), diaminopimelic acid, and Nacetyl-d-glucosamine (NAG) and DAP, which is unique in the cell walls of prokaryotic microorganisms; the teichoic and teichuronic acid are chemically bonded to peptidoglycan [7]. The chemical composition of their cell wall is similar to that of gram-positive bacteria, but because of their well morphological developed and cultural characteristics, actinomycetes have been considered as a group, which separate from the other ordinary bacterial group [8]. Therefore, the current study aimed to screen bacteria Streptomyces isolates in Iraq.

2. Materials and Methods

2.1. Sample collection

Thirty-five samples were collected from different regions of Iraq (Baghdad, Najaf and Babylon), summarized in Table 1. 250 grams of soil samples were collected from regions that were mentioned previously at a depth ranging from 5 to 15 cm, and were kept in polyethylene bags (20 * 40 cm). The soil samples were exposed to the air for a week, also they were pretreated with CaCO3 (with a ratio of 10:1 soil: CaCO3) and kept at ambient temperature for a week, to enrich actinomycetes which usually prefer alkaline conditions and also to reduce the contamination with molds and fungi [9].

No.	Site	Sample	No.	Site	Sample
1101		Abbreviated no.			Abbreviated no.
1	Baghdad	Bag1	19	Najaf	Naj19
2	Baghdad	Bag2	20	Najaf	Naj20
3	Baghdad	Bag3	21	Najaf	Naj21
4	Baghdad	Bag4	22	Najaf	Naj22
5	Baghdad	Bag5	23	Babylon	Bab23
6	Baghdad	Bag6	24	Babylon	Bab24
7	Baghdad	Bag7	25	Babylon	Bab25
8	Baghdad	Bag8	26	Babylon	Bab26
9	Baghdad	Bag9	27	Babylon	Bab27
10	Baghdad	Bag10	28	Babylon	Bab28
11	Baghdad	Bag11	29	Babylon	Bab29
12	Baghdad	Bag12	30	Babylon	Bab30
13	Najaf	Naj13	31	Babylon	Bab31
14	Najaf	Naj14	32	Babylon	Bab32
15	Najaf	Naj15	33	Babylon	Bab33
16	Najaf	Naj16	34	Babylon	Bab34
17	Najaf	Naj17	35	Babylon	Bab35
18	Najaf	Naj18		-	

Table 1. Sites and numbers of soil samples were collected for isolation of locally *Streptomyces* in Iraq

2.2. Isolation and Identification of Streptomyces from Soil

One gram of dried and treated soil samples was used to make suspension, by adding 99 mL of sterile distilled water (stock suspension) and shaking it in a shaker at 160 rpm for 30 minutes at room temperature. Serial dilutions from 0.1-0.001 were made from the stock suspension, and left for 10 minutes. After shaking, 0.1 mL of each dilution was cultured on Yeast Extract and Malt Extract agar (YEME) with Streptomycin 50 ug/mL, then spread by sterile swab for making uniform distribution of the suspension on the surface of the media. The inoculated plates were incubated at 28°C for 7 to 10 days. Based on cultural characteristics, suspected colonies of Streptomyces were selected which are characterized as small, white, pin-point, rough, chalky and a clear zone of inhibition around them. These colonies were confirmed for their identification by types of Gram's stain, aerial and substrate mycelium color, pigment production, and pigment color. These colonies were transferred from the mixed culture into separate agar plates and incubated at 28±1°C for 7 days. In order to obtain a pure growth of *Streptomyces* were re-streaked on International *Streptomyces* project (ISP) to obtain pure colonies used for identification[10-13].

2.3. Streptomyces isolation and identification media

International *Streptomyces* Project (ISP2) Medium of *Streptomyces* isolation (Table2) was used [14]. This medium was prepared by dissolving amount of each component in 1000 mL distilled water and sterilized by autoclaving at 121 C, (15 lb\ inch²) for 15 minutes.

Table 2. Internation	nal <i>Streptomyces</i> Project (ISP2) Medium [17]	

Component	Quantity (g/l)
Yeast extract	4
Malt extract	10
Dextrose	4
	20

2.4. The Best Media Composition for Antimicrobial Production

According to international *Streptomyces* projects (ISP) [14, 15], different media were used for isolation and identification of *Streptomyces* spp. furthermore, to achieving the best types

of media composition for production of antimicrobial metabolites, different media with different composition were used (Table 3). After seven days of incubation, antimicrobial metabolites extraction was carried.

Table 3. International *Streptomyces* projects (ISP) used for isolation and identification of *Streptomyces* [16]

No.	Medial name	Abbreviation
1	Tryptone-yeast extract broth	ISP1
2	Yeast extract-malt extract broth	ISP2
3	Inorganic salts-starch broth	ISP4
4	Glycerol-asparagine broth	ISP5
5	Peptone-Yeast Extract Iron agar	ISP6
6	Tyrosine Agar	ISP7
7	Glycerol yeast extract broth	GYE

2.5. Identification of Bacterial Isolates

Suspected bacterial isolates were primarily identified by Microscopic and cultural examinations, then by the biochemical tests for final identification as follows:

2.5.1. Morphological Characteristics

The morphological characterization of each isolate was first performed by:

2.5.1.1. Colony Characteristics [19]

Bacterial isolates grew on ISP medium were characterized morphologically and physiologically according to the International *Streptomyces* project (ISP).

2.5.1.2. Gram's Stain [20]

A single colony was transferred by a loop to a clean glass slide. The smear was stained with crystal violet, treated with iodine, decolorized by the ethanol (95%), and stained with safranine, then examined by a microscope.

2.5.2. Biochemical properties

Streptomyces spp. isolates selected for biochemical characterization, and various biochemical tests were studied. Many characteristics were studied including czapeck medium [16], sugars utilization medium [17], organic acids formation medium [17], urease test, amylase test, Protease test, cellulase test, Indol production test, Kovae's reagent, phosphatase test, utilization test, gelatinase test and catalase test [18, 19].

2.5.3. Identification onn,..., f bacteria by VITEK 2 system

The VITEK 2 which is recently installed at the Central Health Laboratories/Ministry of Health is an automated microbiology system utilizing growth-based technology. A sterile swab sample was used to transfer a sufficient number of colonies of a pure culture and to suspend them into 3 mL of normal saline (NaCl 0.45%, pH 5-7). Then, turbidity was adjusted by a turbidity meter called the DensiCheck to match 0.5- 0.6 McFarland, which is the proper inoculum density for Gram-negative and gram-positive bacteria as stated by the manufacturer.

3. Results

3.1. Isolation of *Streptomyces* bacteria from the soil

The technique of serial dilution was used to isolate *Streptomyces* bacteria from 35 different samples of soil. Then, the plates were inoculated with soil suspension on media Yeast Extract Malt Extract and the plates were incubated for 7 days with a range of dilution between

0.1-0.001. From the above soil samples on the basis of forming colonies with inhibitory or clear zone of inhibition around them, the suspected *Streptomyces* were obtained, which were small white, chalky and rough.

From 35 soil samples, only 21 samples of soil were suspected to contain *Streptomyces*, mean 60% of them, and 45 strains were obtained with different morphology types per samples of soil. The colonies suspects were selected based on color varied between gray, white and creamy, which were grown on yeast extract malt extract agar.

Also, nine different types of media were tested for their efficiency of *Streptomyces* to support larger number of colonies from the soil (Table 4). These media were used to isolate the Local *Streptomyces* spp. Several types of *Streptomyces* bacteria were isolated on these media, but in this study only one type was used. This bacteria Local *Streptomyces* spp. was isolated from soil collected from Baghdad under depth of 20 cm. The Local *Streptomyces* spp. isolation and purification are shown in Figures 1 and 2.

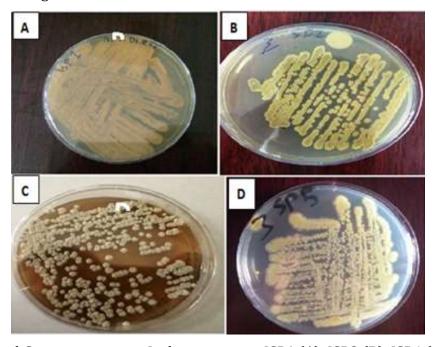


Figure 1. Local *Streptomyces* spp. Isolates grow on ISP1 (A), ISP2 (B), ISP4 (C), and ISP5

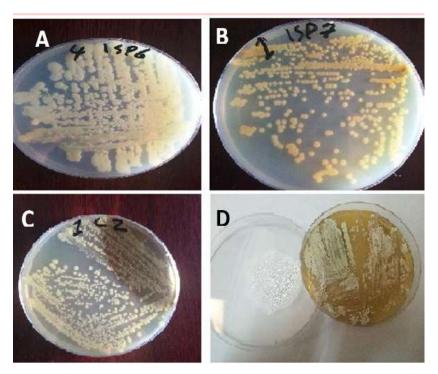


Figure 2. Local *Streptomyces* spp. Isolates grow on ISP6 (A), ISP7 (B), czapeck agar (C), and potato dextrose agar (D)

Table 4. The morphological characteristics of colony on special medium (ISP) and other workers

77 11	6	
Medium	Growth	Aerial mycelium
ISP1	++	Pale yellow
ISP2	++++	Creamy
ISP4	++	Pale white
ISP5	+++	Pale yellow
ISP6	+++	Pale white
ISP7	+++	Creamy
CZ	++	Pale white
PDA	+++	Light white – yellow
N.A.(nutrient agar)	++	Pale - white

+: less, ++: moderate, +++: good, ++++: very good

3.2. Microscopic examination

The microscopic examination of local *Streptomyces* spp. after Gram- staining method was conducted and the observations revealed that local *Streptomyces* is gram positive and rod shaped similar to features of fungal in possessing branched mycelium.

3.3. Biochemical properties

The results of biochemical testes of *Streptomyces* spp. are shown in Table 5. The *Streptomyces* produced extra cellular enzymes like amylase, urease, catalase, protease, Gelatinase, cellulase and phosphatase. Utilization of citrate was positive, with no Melanine reaction production (Medium ISP 2 and Medium ISP6) and Soluble Pigmented (ISP4 and PDA), and negative for indole production.

Reaction	Response	Result
1. Melanine reaction Medium ISP.2	Brownish of medium	Negative
Medium ISP.6	Brownish of medium	Negative
2. Soluble Pigmented	NO Brown	Negative
ISP4 PDA	Dark Brown pigment	Positive
3. Urease	Red to deep pink	Positive
4. Catalase	Bubbles	Positive
5. Amylase	Clear zone	Positive
6.Protease	Clear zone	Positive
7. Gelatinase	Narrow zone	positive
9. Cellulase	Clear zone	Positive
10. Phosphatase	Clear zone	positive
11. Indole production	No color zone	Negative
12. Citrate Utilization	Deep blue color	Positive

Table 5. The biochemical tests properties of *Streptomyces* spp

3.4. Identification of *Streptomyces* by VITEK2 system:

The *Streptomyces* was identified by VITEK 2. By this system of identification, we were able to identify *Streptomyces* accurate. The results of VITEK 2 system agreed with the obtained those of the biochemical tests that were applied for the bacterial isolates (Table 5).

4. Discussion

The technique of serial dilution was used to isolate *Streptomyces* bacteria from 35 different samples of soil. After the plates were inoculated with soil suspension on media Yeast Extract Malt Extract, the plate were incubated for 7 days with a range of dilution between 0.01-0.001, from the above soil samples on the basis of forming colonies with inhibitory or clear zone of inhibition around them the suspected *Streptomyces* were obtained as small white, chalky and rough [20-22].

The results were in agreement with the past study [23]. The colonies size and morphology were in the range from 1 to 10 mm in diameter with relatively smooth surface at the growth beginning, whereas developed to an UN aerial mycelium that appeared as powdery, soft and granular. Isolated Streptomyces colonies were chalky, slowly growing, aerobic, piled, as well as with different color of aerial and substrate mycelium. Also, all colonies that were isolated possessed earthy odors. The colonies were stored in refrigerator at 4°C for further study. The results agreed with those of [24] and [25]. In association with habitats, the Streptomyces diversity exhibited with few different colony types, and process of isolation, where each plate often contained one or few colony types ranging from 2-4 colonies.

Depending on result of [26], they described *Streptomyces* existent in more than one soil types and surface layer of soils are more abundant besides the favoring of alkaline soils, river's mud, compost and riverbeds. On the other hand, the study on *Streptomyces* isolation in the soil [10] showed that physical properties pH, moisture, soil texture, organic matter content and soil reactions were considered as the important factors affecting *Streptomyces* distribution.

A study showed [27] the sample was not collected from surface of soil because of the fact that Streptomyces was found on lower soil surface than 11-15 cm depth into the soil, which may be attributed to the favorable combination of suitable pH and water content. The number of Streptomyces in black-alkaline sandy soil was very high and the second cause of selecting one isolate was the isolation of *Streptomyces* from the soil in complicated characteristic slow growth relative to that of other bacteria. This has resulted in the development of selective isolation [28].

Local Streptomyces is gram positive and rod shaped similar to features of fungal in possessing branched mycelium in their morphology of cell [29]. The Gram-staining response of bacteria is an important criterion; it is different in the ultra-structure and chemical composition of the two main kinds of prokaryotic cells that are found in nature, which is gram positive and gram negative. These two types of cells are different from each other depending on the absence or presence of an outer lipid membrane that is more fundamental and reliable features for the cell of bacteria [30]. All bacteria of gram- positive are bounded by only a single unit lipid membrane and they contain, in general, a thick layer (20-80 nm) of peptidoglycan responsible for gram- stain positive (purple) the retaining [31]. We further characterized **Streptomyces** the strains spp biochemical properties. nutritional uptake and all the isolates yielded similar results obtained by several investigators [4].

The identification of the *Streptomyces* is a very complex process. The *Streptomyces* classification system is mainly dependent on characteristics like the form of spores, melanoma and use of carbon [32]. Morphological and biochemical characteristics are two

important aspects for the classification in the Streptomycetaceae family [33]. By studying the morphological, cultural and biochemical characteristics, observed that the local *Streptomyces* spp. is belonged to the genus of Streptomyces [34], referred to the probability and confidence of identification of Vitek 2 system as the accuracy of the VITEK2 system. It has been pointed out [35] that the VITEK2 system is efficient to handle systems that provide faster results during 4 to 15 h and have reasonably accurate means for the identification of the species of bacterial. One of the mean advantages of the VITEK2 system is the reducing the handling time significantly, having a positive impact on the work flow of the laboratory of clinical microbiology.

5. Conclusions

The identification of the *Streptomyces* is a very complex process. Morphological and biochemical characteristics are two important aspects for the classification in the Streptomycetaceae family. By studying the morphological, cultural and biochemical characteristics, it is observed that the local *Streptomyces* spp. is belonged to the genus of *Streptomyces*.

Authors' contributions

M.H.R., designed this study, B.Q. collected and analyzed the data. B.Q. supervised the collection of samples and the identification. M.H.R. and B.Q., proceeded to the data quality control and the manuscript drafting. . M.H.R. revised the final version.

Consent for publications

All authors agree to have read the manuscript and authorize the publication of the final version of the manuscript.

Conflict declaration

The authors declare that there is no conflict.

Conflict of interest

None of the authors has any conflict of interest to declare.

Availability of data and material

Data are available on request from the authors.

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Ethics approval and consent to participate

Samples collection were obtained from soil.

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References

- 1. Anderson A S, Wellington E. (2001). The taxonomy of *Streptomyces* and related genera. *International Journal of Systematic and Evolutionary Microbiology*, 51(3): 797-814. https://doi.org/10.1099/00207713-51-3-797
- 2. Labeda D P. (2011). Multilocus sequence analysis of phytopathogenic species of the genus *Streptomyces*. *International Journal of Systematic and Evolutionary Microbiology*, 61(10): 2525-2531. https://doi.org/10.1099/ijs.0.028514-0
- 3. Qasim B, Risan M H. (2017). Antitumor and Antimicrobial Activity of Antibiotic Produced by *Streptomyces* spp. *World Journal of Pharmaceutical Research*, 6(4): 116-128.
- 4. Shobha K, Onkarappa R, Goutham S, Raghavendra H. (2012). Screening

- biological activities of a *Streptomyces* species isolated from soil of Agumbe, Karnataka, India. *Int J Dug Dev Res*, 4(3): 104-114.
- 5. Yun T, Zhang M, Zhou D, Jing T, Zang X, Qi D, Chen Y, Li K, Zhao Y, Tang W. (2021). Anti-Foc RT4 Activity of a Newly Isolated *Streptomyces* sp. 5–10 From a Medicinal Plant (*Curculigo capitulata*). *Frontiers in Microbiology*, 11: 3544. https://doi.org/10.3389/fmicb.2020.6 10698
- 6. Mahon C R, Lehman D C, Manuselis G. (2018). Textbook of diagnostic microbiology-e-book: *Elsevier Health Sciences*. 1088 pages,
- 7. Davenport R J, Curtis T P, Goodfellow M, Stainsby F M, Bingley M. (2000). Quantitative use of fluorescent in situ hybridization to examine relationships between mycolic acid-containing actinomycetes and foaming in activated sludge plants. *Applied and environmental Microbiology*, 66(3): 1158-1166. https://doi.org/10.1128/AEM.66.3.11 58-1166.2000
- 8. Das S, Lyla P, Khan S A. (2008). Distribution and generic composition of culturable marine actinomycetes from the sediments of Indian continental slope of Bay of Bengal. *Chinese Journal of Oceanology and Limnology*, 26(2): 166-177. https://doi.org/10.1007/s00343-008-0166-5
- 9. T Abdulhameed Z. (2013). The isolation and study of morphological characterization of *Streptomyces* isolated from the soil as a source of active antibiotic. *College Of Basic Education Researches Journal*, 12(3): 745-752.
- 10. Nonoh J O, Lw W, Masiga D, Herrmann R, Presnail J K, Schepers E, Okech M A, Bagine R, Mungai P, Nyende A B. (2010). Isolation and characterization of *Streptomyces*

- species with antifungal activity from selected national parks in Kenya. *African Journal of Microbiology Research*, 4(9): 856-864. https://doi.org/10.5897/AJMR.90004
- 11. Ali-Soufi M. Shahriari A. Shirmohammadi E. Fazeli-Nasab B. (2019). Investigation of Dust Microbial Community and Identification of its Dominance Species in Northern Regions of Sistan and Baluchestan Province. Journal of Water and Soil Science (Science and Technology of Agriculture and Natural Resources), 23(1): 309-320. https://doi.org/10.29252/jstnar.23.1.
- 12. Abbasi-Moghadam J, Shahriari A, Fazeli-Nasab B. (2017). Investigation of bacteria and fungi populations associated with airborne dust during "wind of 120 days" blowing in the urban areas of Sistan plain. Paper presented at the 15th Iranian Soil Science Congress, Isfahan University of Technology, Isfahan, Iran, Congress COI: SSCI15, Article COI: SSCI15_687.
- 13. Ali-Soufi M. Shahriari A. Shirmohammadi E, Fazeli-Nasab B. (2017). Identification and isolation of microorganisms associated with airborne dust loaded over Sistan plain. Paper presented at the 15th Iranian Soil Science Congress, Isfahan University of Technology, Isfahan, Iran, Congress COI: SSCI15, Article COI: SSCI15_895.
- 14. Lapaz M, Huguet-Tapia J, Siri M, Verdier E, Loria R, Pianzzola M. (2017). Genotypic and phenotypic characterization of *Streptomyces* species causing potato common scab in Uruguay. *Plant Disease*, 101(8): 1362-1372. https://doi.org/10.1094/PDIS-09-16-1348-RE
- 15. Wei Z, Xu C, Wang J, Lu F, Bie X, Lu Z. (2017). Identification and

- characterization of *Streptomyces* flavogriseus NJ-4 as a novel producer of actinomycin D and holomycin. *PeerJ*, 5: e3601. PubMed: 28740758
- Shariffah-Muzaimah S. Idris Dzolkhifli Madihah A. 0. Kamaruzzaman S, Maizatul-Suriza M. (2018).Characterization of Streptomyces spp. isolated from the rhizosphere of oil palm and evaluation of their ability to suppress basal stem rot disease in oil palm seedlings when applied as powder formulations in a glasshouse trial. World Journal of *Microbiology and Biotechnology*, 34(1):
 - https://doi.org/10.1007/s11274-017-2396-1
- 17. Yu Z, Han C, Yu B, Zhao J, Yan Y, Huang S, Liu C, Xiang W. (2020). Taxonomic characterization, and secondary metabolite analysis of *Streptomyces* triticiradicis sp. nov.: A novel actinomycete with antifungal activity. *Microorganisms*, 8(1): 77. https://doi.org/10.3390/microorganisms8010077
- 18. Church D L. (2016). Biochemical tests for the identification of aerobic bacteria Clinical Microbiology Procedures Handbook, Fourth Edition (pp. 3.17. 11.11-13.17. 48.13): American Society of Microbiology.
- 19. Odds F. (1981). Biochemical tests for identification of medical bacteria. *Journal of Clinical Pathology*, 34(5): 572. PMCID: PMC493360
- 20. Ramazani A, Moradi S, Sorouri R, Javani S, Garshasbi M. (2013). Screening for antibacterial activity of *Streptomyces* species isolated from Zanjan province, Iran. *Int J Pharm Chem Biol Sci*, 3(2): 342-349.
- 21. Ali-Soufi M, Shahriari A, Shirmohammadi E, Fazeli-Nasab B. (2017). Seasonal changes biological characteristics of airborne dust in Sistan plain, Eastern Iran. Paper presented at the International

- Conference on Loess Research, Gorgan University of Agricultural Sciences and Natural Resources, Gorgan, Iran.
- 22. Ali-Soufi M, Shahriari A, Shirmohammadi E, Fazeli-Nasab B. (2017). Investigation of biological properties and microorganism identification in susceptible areas to wind erosion in Hamoun wetlands. Paper presented at the Congress on restoration policies and approaches of Hamoun international wetland Zabol
- 23. Saadoun I, Al-Joubori B, Al-Khoury R. (2015). Testing of production of inhibitory bioactive compounds by soil *Streptomycetes* as preliminary screening programs in UAE for anticancer and anti-bacterial drugs. *Int. J. Curr. Microbiol. App. Sci*, 4(3): 446-459.
- 24. Zhou J-P, Gu Y-Q, Zou C-S, Mo M-H. (2007). Phylogenetic diversity of bacteria in an earth-cave in Guizhou Province, Southwest of China. *Journal of microbiology*, 45(2): 105-112.
- 25. Portillo M C, Saiz-Jimenez C, Gonzalez M. (2009).Molecular characterization of total and metabolically active bacterial communities of "white colonizations" in the Altamira Cave, Spain. Research microbiology. 160(1): 41-47. https://doi.org/10.1016/j.resmic.200 8.10.002
- 26. Kariminik A, Baniasadi F. (2010). Pageantagonistic activity of Actinomycetes on some Gram negative and Gram positive bacteria. *World Applied Sciences Journal*, 8(7): 828-832.
- 27. Augustine S, Bhavsar S, Kapadnis B. (2005). A non-polyene antifungal antibiotic from *Streptomyces* albidoflavus PU 23. *Journal of Biosciences*, 30(2): 201-211. https://doi.org/10.1007/BF02703700
- 28. El-Naggar M Y, El-Assar S A, Abdul-Gawad S M. (2006). Meroparamycin production by newly isolated

- Streptomyces sp. strain MAR01: taxonomy, fermentation, purification and structural elucidation. *Journal of microbiology*, 44(4): 432-438.
- 29. Rana S, Salam M. (2014). Antimicrobial potential of actinomycetes isolated from soil samples of Punjab. *India. J Microbiol Exp*, 1(2): 00010.
- Gupta 30. R S. (1998).Protein phylogenies and signature sequences: evolutionary reappraisal of relationships among archaebacteria, eukaryotes. eubacteria, and Microbiology and Molecular Biology Reviews. 62(4): 1435-1491. https://doi.org/10.1128/MMBR.62.4. 1435-1491.1998
- 31. Gupta R S. (1998). What are archaebacteria: life's third domain or monoderm prokaryotes related to Gram-positive bacteria? Α new proposal for the classification of organisms. prokaryotic Molecular microbiology. 29(3): 695-707. https://doi.org/10.1046/j.1365-2958.1998.00978.x
- 32. Buchanan R, NE G. (1974). Bergey's manual of determinative bacteriology, 7th edn, London. *Bergey Taxon*, 24: 377-378.
- 33. Naine S J, Devi C S, Mohanasrinivasan V. (2014). Antimicrobial, Antioxidant and Cytotoxic Activity of Marine *Streptomyces* parvulus VITJS11 Crude Extract. *Brazilian archives of biology and technology*, 58: 198-207. https://doi.org/10.1590/S1516-8913201400173
- 34. Aziz Z S, Al-Muhanna A S, Salman A J, Alzuhairi M A. (2014). *Klebsiella* and *Raoultella* biotyping and probability of identification by Vitek-2 system. *IJIRSET*, 3(4): 11289-11294.
- 35. Garcia-Garrote F, Cercenado E, Bouza E. (2000). Evaluation of a new system, VITEK 2, for identification and antimicrobial susceptibility testing of *enterococci. Journal of Clinical*

Microbiology, 38(6): 2108-2111. 08-2111.2000 https://doi.org/10.1128/JCM.38.6.21

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